

## The first total synthesis of telephiose A

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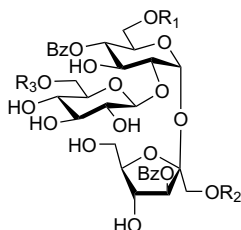
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**Abstract**—The first total synthesis of telephiose A (**1**), a novel trisaccharide ester having two acetyl groups and two benzoyl groups, was achieved by using glucosyl donor **6** and disaccharide acceptor **12**. The crucial key step was the stereoselective construction of the  $\beta$ -D-glucosidic linkage featuring the neighboring group participation of the 2-*O*-*N*-phenylcarbamoyl group (of donor **6**), which can be selectively deprotected in the presence of acetyl and benzoyl groups. Donor **6** was prepared from D-glucose in eight steps (33% yield), whereas acceptor **12** was prepared from sucrose in six steps (35% yield). Precursors **6** and **12** were reacted in subsequent reactions (five steps) to afford **1** in 22% yield.

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Three new oligosaccharide esters (telephiose A–C, Fig. 1) have been isolated from *Polygala telephioides* WILLD, a plant, that is, widely distributed in southern China and employed as a detoxification agent for heroin poisoning in China. The structures of the esters have been recently characterized using spectroscopic studies.<sup>1</sup> Due to our interest in the detoxification activities and the structures of these partially acylated oligosaccharide esters, we decided to undertake the total synthesis of telephiose A (**1**) via a key step in the stereoselective construction of the  $\beta$ -D-glucosidic linkage via neighboring group participation of 2-*O*-acyl type protecting group,



Telephiose A (**1**) : R<sub>1</sub> = R<sub>2</sub> = Ac, R<sub>3</sub> = H  
 B : R<sub>1</sub> = H, R<sub>2</sub> = R<sub>3</sub> = Ac  
 C : R<sub>1</sub> = R<sub>2</sub> = R<sub>3</sub> = H

**Figure 1.** The structures of telephioses.

**Keywords:** Telephiose; Total synthesis; Oligosaccharide esters; *N*-Phenylcarbamoyl group.

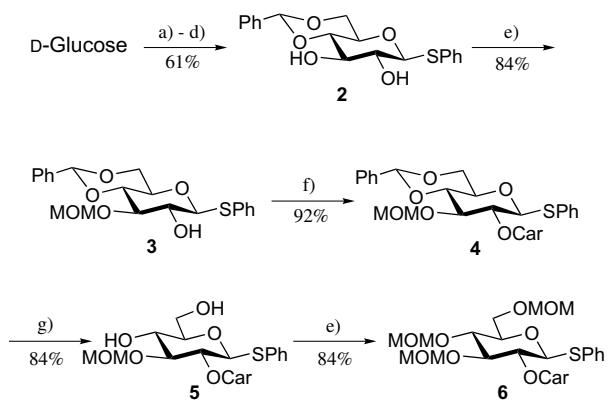
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which could be deprotected in the presence of acyl groups of the glucosyl donor.

In a previous Letter,<sup>2</sup> we reported on the construction of four types of glycosidic linkages via a universal glucosyl donor that involves the neighboring group participation of an *N*-phenylcarbamoyl (Car) group and the S<sub>N</sub>2 displacement reaction at C-2. The Car group is stable from pH 1 to 12 in aqueous solutions,<sup>3</sup> and because of the difficulty in the deprotection, the Car group has yet to be widely employed for the protection of hydroxyl groups. To the best of our knowledge, the Car protecting group has not been employed in the synthesis of complex natural products, which often require delicate chemical differentiation of various protecting groups under mild conditions. We have successfully developed a novel deprotection procedure that does not affect acyl, silyl, methoxymethyl, benzylidene acetal, and isopropylidene acetal protecting groups,<sup>4</sup> and therefore, has allowed the Car group to become a valuable tool in natural products syntheses.

Herein, we describe the practical synthesis of telephiose A (**1**) from D-glucose and D-sucrose featuring the properties of the Car group.

As shown in Scheme 1, glucosyl donor **6** was prepared from D-glucose in eight steps. Initially, phenyl 4,6-*O*-benzylidene-1-thio- $\beta$ -D-glucopyranoside (**2**)<sup>5</sup> was prepared from D-glucose in four steps (61% yield). Treatment of **2** with methoxymethyl chloride (MOM-Cl,

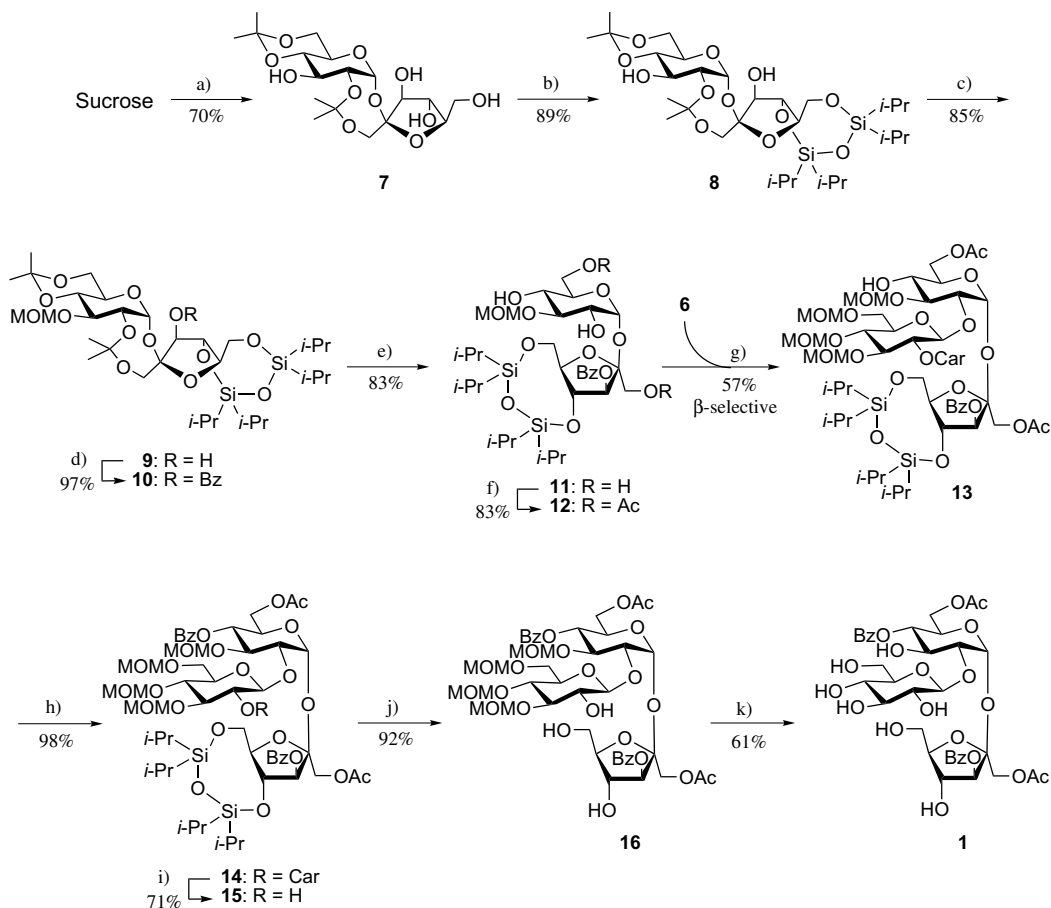


**Scheme 1.**<sup>9</sup> Reagents and conditions: (a)  $\text{Ac}_2\text{O}$ ,  $\text{AcONa}$ ; (b)  $\text{PhSH}$ ,  $\text{BF}_3 \cdot \text{Et}_2\text{O}/(\text{ClCH}_2)_2$ ; (c)  $\text{NaOMe}/\text{MeOH}$ ; (d)  $\text{PhCH}(\text{OCH}_3)_2$ ,  $p\text{-TsOH}/\text{DMF}$ ; (e)  $\text{MOM-Cl}$ ,  $i\text{-Pr}_2\text{NEt}/\text{CH}_2\text{Cl}_2$ ; (f)  $\text{Ph-NCO}/\text{Py}$ ; (g) 70% aq  $\text{AcOH}$ .

1.5 equiv) and *N*-ethyl-diisopropylamine (1.2 equiv) in  $\text{CH}_2\text{Cl}_2$  at  $0^\circ\text{C}$  for 12 h afforded **3** (84% yield), which was reacted with phenyl isocyanate (2.0 equiv) in pyridine with stirring for 5 h at rt. Upon the addition of  $\text{MeOH}$ , the reaction mixture was removed by evaporation, and the crude product was recrystallized to give

the corresponding 2-*O*-Car derivative **4** in 92% yield. Acid hydrolysis of **4** using 70% aq  $\text{AcOH}$  at  $60^\circ\text{C}$  for 1 h gave 4,6-diol derivative **5**, which was then treated with  $\text{MOM-Cl}$  (2.5 equiv) and *N*-ethyl-diisopropylamine (2.5 equiv) in  $\text{CH}_2\text{Cl}_2$  for 4 h to afford glucosyl donor **6**<sup>6</sup> in 71% yield (two steps).

On the other hand, as showed in **Scheme 2**, acceptor **12** was prepared from sucrose in six steps. Following Manzo's procedure,<sup>7</sup> sucrose, acetone dimethylacetal (10 equiv), and cerium(IV) ammonium nitrate (0.2 equiv) were reacted to afford 2,1':4,6-di-*O*-isopropylidene-sucrose (**7**) in 70% yield. The selective protection<sup>8</sup> of the 4',6'-OH groups of **7** involved the addition of 1,3-dichloro-1,1,3,3-tetraisopropylidisiloxane (1.2 equiv) (in three portions) to a mixture of **7** and imidazole (1.0 equiv) in pyridine at  $0^\circ\text{C}$ . After maintaining the reaction mixture for 8 h, and upon the disappearance of **7** (determined by TLC),  $\text{MeOH}$  was added to the reaction mixture. Typical work up procedures, followed by purification using silica gel column chromatography afforded **8** in 89% yield. Purified **8** was reacted with  $\text{MOM-Cl}$  (1.2 equiv) and *N*-ethyl-diisopropylamine (1.2 equiv) in  $\text{CH}_2\text{Cl}_2$  at rt for 6 h to give 3-*O*-methoxymethyl derivative **9** (85% yield), which was treated with benzoyl chloride (2.0 equiv) in pyridine to afford



**Scheme 2.**<sup>9</sup> Reagents and conditions: (a)  $\text{Ce}(\text{N}_2\text{H}_4)_2(\text{NO}_3)_6$ ,  $(\text{CH}_3)_2\text{C}(\text{OCH}_3)_2/\text{DMF}$ ; (b)  $[(\text{CH}_3)_2\text{CH}_2\text{SiCl}]_2\text{O}-[\text{ClSi}[\text{CH}(\text{CH}_3)_2]_2]$ , imidazole/ $\text{DMF}$ ; (c)  $\text{MOM-Cl}$ ,  $i\text{-Pr}_2\text{NEt}/\text{CH}_2\text{Cl}_2$ ; (d)  $\text{BzCl}$ ,  $\text{Py}$ ; (e) 0.1 M  $\text{HCl-MeOH}$ ; (f)  $\text{AcCl}$ ,  $\text{Py}$ ; (g)  $\text{NIS}$ ,  $\text{TMSOTf}$ ,  $\text{MS4A}/\text{CH}_2\text{Cl}_2$ ; (h)  $\text{BzCl}$ ,  $\text{Py}$ ; (i)  $\text{Bu}_4\text{NNO}_2/\text{DMF}$ ; (j)  $\text{TBAF}/\text{THF}$ ; (k) 90%  $\text{AcOH}$  aq.

the corresponding 3'-*O*-benzoate derivative as an oil (97% yield); the structure of **10** was confirmed using <sup>1</sup>H NMR spectroscopy [downfield shift of H-3'(fruc) and elemental analysis. Direct mono-benzoylation of **10** gave 3-*O*-benzoate (glc) instead 3'-*O*-benzoyl (glc) derivative, by the way. Selective hydrolysis of **10** at rt using 0.1 M HCl–MeOH, followed by neutralization using Dowex (OH<sup>-</sup> form), afforded de-*O*-isopropylidene **11** (83% yield), which was treated with acetyl chloride (1.5 equiv) in pyridine at 0 °C for 30 min to afford 6,1'-di-*O*-acetate **12** as an oil<sup>6</sup> (83% yield). The structure of **12** was confirmed using <sup>1</sup>H NMR spectroscopy [downfield shift of H-6 and H-1'] and elemental analysis.

Using donor **6** and disaccharide acceptor **12**, telephiose A (**1**) was synthesized as follows: to a stirred mixture of **6** (1.0 equiv), **12** (0.9 equiv), and MS 4A in CH<sub>2</sub>Cl<sub>2</sub> (after stirring for 30 min under argon at rt) was added (dropwise) dry *N*-iodosucinimide (1.5 equiv), followed by trimethylsilyl trifluoromethanesulfonate (TMSOTf, 0.3 equiv) at -40 °C. Upon disappearance of the starting material using TLC (1 h), the reaction mixture was neutralized with Et<sub>3</sub>N, filtered, diluted with CHCl<sub>3</sub>, and washed with aq NaHCO<sub>3</sub>, aq Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>, brine, and water. The organic layer was dried over anhydrous MgSO<sub>4</sub>, removed by evaporation, then purified using silica gel column chromatography to give trisaccharide **13** as an oil [<sup>1</sup>H NMR, *J*<sub>1,2</sub> = 9.7 Hz, H-1 (β-linkage)] (57% yield). In addition to the main product, the formation of a small amount of a side product, the 4-*O*-glucoside (10%), was observed. Benzoylation of the 4-OH group of **13** with benzoyl chloride (1.5 equiv) in pyridine gave **14** (98% yield), which was confirmed using <sup>1</sup>H NMR spectroscopy [down field shift of H-4 (glc)] and elemental analysis. The 2-*O*-*N*-phenylcarbamoyl group of **14** was removed using Bu<sub>4</sub>NNO<sub>2</sub> (3.0 equiv) in DMF at 90 °C for 12 h under argon to give **15** as an oil (71% yield), which was treated with Bu<sub>4</sub>NF (TBAF) in THF to afford de-*O*-silylated **16** as an oil (92% yield). Deprotection conditions of the MOM groups using 90% aq AcOH at rt resulted in a mixture of **1** (yield 61%) and a side product (cleavage of the fructosyl linkage, ca. 20%). Purification using silica gel column chromatography (CHCl<sub>3</sub>–MeOH) afforded pure **1**; the optical rotation values and <sup>1</sup>H NMR data are in good agreement with those reported.<sup>1,6</sup>

Herein, we described the first total synthesis of telephiose A using a glucosyl donor (based on D-glucose) and a disaccharide acceptor (based on sucrose). Our synthetic methodology illustrates the usefulness of a Car protecting group in the synthesis of oligosaccharide esters.

#### Acknowledgments

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#### Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2007.03.138.

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- IR (KBr, neat), <sup>1</sup>H NMR, and other physical data of compound **6**, **12**, and **1**.  
Compound **6**: mp 173.5–174.0 °C (hexane–EtOH); [ $\alpha$ ]<sub>D</sub><sup>25</sup> +20.8 (*c* 0.7, CHCl<sub>3</sub>);  $\nu_{\text{NH}}$  2928 cm<sup>-1</sup>,  $\nu_{\text{C=O}}$  1738 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.54–7.05 (5H, m, Ph), 6.83 (1H, s, -NH-), 5.15 (1H, dd, *J*<sub>2,1</sub> = 9.7 Hz, *J*<sub>2,3</sub> = 9.2 Hz, H-2), 4.82, 4.72 (2H, each d, *J*<sub>AB</sub> = 7.4 Hz, CH<sub>2</sub>OCH<sub>3</sub>), 4.74, 4.68 (2H, each d, *J*<sub>AB</sub> = 7.8 Hz, CH<sub>2</sub>OCH<sub>3</sub>), 4.72, 4.67 (2H, each d, *J*<sub>AB</sub> = 6.2 Hz, CH<sub>2</sub>OCH<sub>3</sub>), 4.68 (1H, d, H-1), 3.84 (1H, dd, *J*<sub>6a,5</sub> = 2.2 Hz, *J*<sub>6a,6b</sub> = 11.5 Hz, H-6a), 3.73 (1H, dd, *J*<sub>6b,5</sub> = 5.1 Hz, H-6b), 3.68 (1H, dd, *J*<sub>3,4</sub> = 9.7 Hz, H-5), 3.57 (1H, dd, *J*<sub>4,5</sub> = 9.2 Hz, H-4), 3.41 (1H, ddd, H-5), 3.36, 3.33, 3.30 (9H, each s, OCH<sub>3</sub>). Anal. Calcd for C<sub>25</sub>H<sub>33</sub>NO<sub>9</sub>S (523.60): C, 67.86; H, 5.34; N, 2.39. Found: C, 67.91; H, 5.54; N, 2.37.  
Compound **12**: [ $\alpha$ ]<sub>D</sub><sup>26</sup> -15.0 (*c* 0.7, CHCl<sub>3</sub>); IR (KBr, neat)  $\nu_{\text{C=O}}$  1732 cm<sup>-1</sup>, 1738 cm<sup>-1</sup>, 1746 cm<sup>-1</sup>,  $\nu_{\text{OH}}$  3455 cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.08–7.27 (5H, m, Ph), 5.61 (1H, d, *J*<sub>3',4'</sub> = 8.6 Hz, H-3'), 5.52 (1H, d, *J*<sub>1,2</sub> = 3.4 Hz, H-1), 4.76 (1H, dd, *J*<sub>6a,5</sub> = 8.6 Hz, *J*<sub>6a,6b</sub> = 8.3 Hz, H-6a), 4.71, 4.58 (2H, each d, *J*<sub>AB</sub> = 6.9 Hz, CH<sub>2</sub>OCH<sub>3</sub>), 4.43 (1H, d, *J*<sub>1a',1b'</sub> = 12.0 Hz, H-1a'), 4.32 (1H, d, H-1b'), 4.31 (1H, dd, *J*<sub>6b,5</sub> = 9.5 Hz, H-6b), 4.06–4.05 (2H, m, H-4', H-5'), 3.94–3.90 (2H, m, H-6a', H-3), 3.56–3.50 (2H, m, H-5, H-6b'), 3.41 (3H, s, OCH<sub>3</sub>), 3.39–3.32 (2H, m, H-2, H-4), 2.08 (6H, s, COCH<sub>3</sub> × 2), 1.12–0.95 (24H, m, TIPDS); Anal. Calcd for C<sub>37</sub>H<sub>60</sub>O<sub>16</sub>Si<sub>2</sub> (817.03): C, 54.39; H, 7.40. Found: C, 54.51; H, 7.64.  
Telephiose (**1**): [Synthetic]: colorless syrup; [ $\alpha$ ]<sub>D</sub><sup>24</sup> -14.8 (*c* 0.36, MeOH) <sup>1</sup>H NMR (600 MHz, Pyridine-*d*<sub>5</sub>)  $\delta$  8.31–7.41 (10H, m, Ph), 6.30 (1H, d, *J*<sub>1,2</sub> = 3.9 Hz, H-1 (Glc1)), 6.28 (1H, d, *J*<sub>3,4</sub> = 8.9 Hz, H-3 (Fruc)), 5.69 (1H, dd, *J*<sub>4,3</sub> = 9.9 Hz, *J*<sub>4,5</sub> = 9.9 Hz, H-4 (Glc1)), 5.17 (1H, dd, *J*<sub>4,5</sub> = 8.0 Hz, H-4 (Fruc)), 5.14 (1H, d, *J*<sub>2,3</sub> = 12.2 Hz, H-2 (Fruc)), 5.09 (1H, d, *J*<sub>1,2</sub> = 8.0 Hz, H-1 (Glc2)), 4.87 (1H, m, H-5 (Glc1)), 4.69 (1H, m, H-5 (Fruc)), 4.62 (1H, m, *J*<sub>1,2</sub> = 12.2 Hz, H-1 (Fruc)), 4.59 (1H, dd, *J*<sub>3,2</sub> = 9.9 Hz, H-3 (Glc1)), 4.54–4.48 (3H, m, H-6 (Glc1), H-6' (Glc1), H-6 (Glc2)), 4.47 (1H, dd, *J*<sub>5,6</sub> = 6.4 Hz, *J*<sub>6,6'</sub> = 12.3 Hz, H-6 (Fruc)), 4.38 (1H, dd, *J*<sub>5,6</sub> = 3.2 Hz, H-6' (Fruc)), 4.30 (1H, dd, *J*<sub>5,6'</sub> = 5.8 Hz, *J*<sub>6',6</sub> = 12.1 Hz, H-6' (Glc2)), 4.17–3.98 (4H, m, H-4 (Glc2), H-3 (Glc2), H-2 (Glc2), H-2 (Glc1)), 3.93 (1H, m, H-5 (Glc2)), 2.10, 1.91 (6H, each s, COCH<sub>3</sub> × 2).  
[Natural]: syrup; [ $\alpha$ ]<sub>D</sub><sup>24</sup> -11.0 (*c* 0.45, MeOH) <sup>1</sup>H NMR (300 MHz, Pyridine-*d*<sub>5</sub>)  $\delta$  8.26–7.40 (10H, m, Ph), 6.31 (1H,

d,  $J_{1,2} = 3.7$  Hz, H-1 (Glc1)), 6.30 (1H, d,  $J_{3,4} = 8.6$  Hz, H-3 (Fruc)), 5.67 (1H, dd,  $J_{4,3} = 9.8$  Hz,  $J_{4,5} = 9.8$  Hz, H-4 (Glc1)), 5.15 (1H, dd,  $J_{4,5} = 7.9$  Hz, H-4 (Fruc)), 5.12 (1H, d,  $J_{2,3} = 12.2$  Hz, H-2 (Fruc)), 5.08 (1H, d,  $J_{1,2} = 7.9$  Hz, H-1 (Glc2)), 4.87 (1H, m, H-5 (Glc1)), 4.68 (1H, m, H-5 (Fruc)), 4.60 (1H, m,  $J_{1,2} = 12.2$  Hz, H-1 (Fruc)), 4.59 (1H, dd,  $J_{3,2} = 9.8$  Hz, H-3 (Glc1)), 4.51–4.50 (3H, m, H-6 (Glc1), H-6' (Glc1), H-6 (Glc2)), 4.43 (1H, dd,  $J_{5,6} = 6.4$  Hz,  $J_{6,6'} = 12.4$  Hz, H-6 (Fruc)), 4.37 (1H, dd,  $J_{5,6} = 3.4$  Hz, H-6' (Fruc)), 4.31 (1H, dd,  $J_{5,6'} = 5.8$  Hz,

$J_{6',6} = 11.9$  Hz, H-6' (Glc2)), 4.13–4.03 (4H, m, H-4 (Glc2), H-3 (Glc2), H-2 (Glc2), H-2 (Glc1)), 3.91 (1H, m, H-5 (Glc2)), 2.07, 1.89 (6H, each s,  $\text{COCH}_3 \times 2$ ).

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9. For experimental procedure and their physical data of compounds **3–5**, **7–11**, and **13–16**, see Supplementary data at doi:10.1016/j.tetlet.2007.03.138.